

# SPINeasy DNA Kit for Soil

Cat. No.: 116530050 (50 preps) & 116530000 (5 preps)



## Quick-Start Protocol

Revision 1.0 (May 2020)

### Notes before starting:

- Add 35 mL (3.5 mL for sample kit) absolute ethanol to Binding Buffer S and mark on the bottle.
- Add 50 mL (5 mL for sample kit) absolute ethanol to Wash Buffer S and mark on the bottle.
- Prepare 100x2.0mL ( 10x2.0mL for sample kit ) microcentrifuge tubes.
- Vortex the sample at full speed for 10 min if a FastPrep<sup>®</sup> Instrument is unavailable. Secure samples on the vortex through an adapter to ensure homogenization.
- Centrifugation speed stated in the manual will be a guideline, use the maximum speed available if 14,000 x g is not feasible.

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| Lyse   | <ol style="list-style-type: none"><li>1 Add 100–500 mg soil sample to <b>Lysing Matrix E</b> tube.</li><li>2 Add 980 <math>\mu</math>L <b>Lysis Buffer S1</b>, 120 <math>\mu</math>L <b>Lysis Buffer S2</b> and 10 <math>\mu</math>L <b>RNase A Solution</b> to sample, vortex to mix.</li><li>3 Homogenize in a FastPrep Instrument for 20 seconds at speed setting of 6.0 m/s.</li><li>4 Centrifuge at 14,000 x g for 5 min.</li></ol> |
| Purify | <ol style="list-style-type: none"><li>5 Carefully transfer the supernatant to a clean 2.0 mL microcentrifuge tube (self–provided).</li><li>6 Add 250 <math>\mu</math>L <b>Inhibitor Removal S</b> to the transferred supernatant and shake 10 times to mix.</li><li>7 Centrifuge at 14,000 x g for 10 min.</li><li>8 Transfer 900 <math>\mu</math>L supernatant to a clean 2.0 mL microcentrifuge tube (self–provided).</li></ol>        |
| Bind   | <ol style="list-style-type: none"><li>9 Add 900 <math>\mu</math>L of <b>Binding Buffer S</b> to the transferred supernatant.</li><li>10 Transfer 800 <math>\mu</math>L of mixture to <b>Column S1</b> placed on top of a <b>2.0 mL Collection Tube</b> (provided).</li><li>11 Centrifuge at 14,000 x g for 1 min. Empty collection tube and reuse. Repeat the process once and discard remaining mixture.</li></ol>                      |
| Wash   | <ol style="list-style-type: none"><li>12 Add 500 <math>\mu</math>L of <b>Wash Buffer S</b> to the column. Centrifuge at 14,000 x g for 1 min. Empty collection tube and reuse. Repeat the washing step.</li><li>13 Without addition of any liquid, centrifuge at 14,000 x g for 2 min to dry the column.</li></ol>   |
| Dry    | <ol style="list-style-type: none"><li>14 Discard collection tube and place the column into a <b>1.5 mL Collection Tube</b> (provided).</li><li>15 Air dry the column for 5 min at room temperature.</li><li>16 Heat <b>DES Buffer</b> to 55 <math>^{\circ}</math>C using a water bath while waiting.</li></ol>   |
| Elute  | <ol style="list-style-type: none"><li>17 Add 100 <math>\mu</math>L of pre–heated <b>DES Buffer</b> to the center of column.</li><li>18 Centrifuge at 14,000 x g for 1 min to elute DNA.</li><li>19 Eluted DNA is now ready for downstream applications. Store at –20 <math>^{\circ}</math>C for extended periods or 4 <math>^{\circ}</math>C until use.</li></ol>  |



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